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## STUDIES ON MARINE NATURAL PRODUCTS. IV THE STEREOCHEMISTRY OF 13-MEMBERED CARBOCYCLIC CEMBRANOLIDE DITERPENES FROM THE SOFT CORAL LOBOPHYTUM PAUCIFLORUM (EHRENBERG)

Yasuji Yamada\*, Sukeji Suzuki and Kazuo Iguchi Tokyo College of Pharmacy, Hachioji, Tokyo 192-03, Japan

Hiroyuki Kikuchi, Yasumasa Tsukitani, Haruo Horiai and Fumio Shibayama Tokyo Research Laboratories, Fujisawa Pharmaceutical Co., Ltd., Nukuikitamachi, Koganei, Tokyo 184, Japan

<u>Summary</u>: The relative and absolute stereochemistry of two 13-membered carbocyclic cembranolides was determined as shown in  $\underline{1}$  and  $\underline{2}$  on the basis of NMR spectral evidence.

A number of marine cembranolide diterpenes have been recently found in soft  $coral_{,}^{1)}$  and in some cases these diterpenes have been reported to have interesting biological properties,<sup>2)</sup> such as an anticancer activity. Most of these diterpenes possess a 14-membered carbocyclic ring system, to which a X- or 5-lactone moiety is attached in many cases. We have already described the isolation and the planar structures of two cembranolide diterpenes, (1) and (2) from the Japanese soft coral <u>Lobophytum pauciflorum</u> (Ehrenberg)(Coelenterata, Anthozoa, Alcyonaria, Alcyonacea), each of which contains a unique 13-membered carbocyclic ring system. The stereochemistry of two trisubstituted carbon-carbon double bonds have been assigned as E-type geometry from the <sup>13</sup>C-NMR chemical shifts of the olefin methyl carbons. We now wish to describe evidence for the stereochemistry of the chiral centers at C-3a, 5, 6 and 14a as depicted in <u>1</u> and <u>2</u>.

The absolute configuration of the secondary hydroxyl group at C-5 in (2) we determined by applying a <sup>1</sup>H-NMR lanthanide induced shifts(LIS) method<sup>4)</sup> and Mosher's <sup>19</sup>F-NMR configuration-correlation method<sup>5)</sup> for the diastereomeric  $\alpha$ -methoxy- $\alpha$ -trifluoromethylphenylacetyl(MTPA) esters. Acylation of (2) with <u>R</u>-(+)-



and  $\underline{S}$ -(-)-MTPA chlorides gave the diastereomeric esters (<u>3</u>) and (<u>4</u>), respectively. The LIS values of OMe and <sup>19</sup>F-NMR chemical shifts of CF<sub>3</sub> in the pair of the diastereomeric esters are shown in Table I. The negative sign of the  $\triangle$ LIS<sub>OME</sub> value<sup>4</sup>) for MTPA esters suggests that the secondary hydroxyl group at C-5 in <u>2</u> has the <u>S</u> configuration. In addition, Mosher's model<sup>5</sup>) predicts that the <u>S</u>-(-)-MTPA ester (<u>4</u>) will show CF<sub>3</sub> resonance downfield relative to the same resonance in the <u>R</u>-(+)-MTPA ester (<u>3</u>) if the secondary hydroxyl group has the <u>S</u> configuration. Alternatively, in the case of the <u>R</u> configuration, the prediction will be reversed. The data on the CF<sub>3</sub> resonance in Table I revealed that the secondary hydroxyl group at C-5 has the <u>S</u> configuration. Application of Horeau's method<sup>6</sup>) to the secondary alcohol (<u>2</u>) also showed the configuration at C-5 to be <u>S</u> : observed rotation, (<u>a</u>)<sup>24°</sup><sub>D</sub> -1.1°(c 2.54, C<sub>6</sub>H<sub>6</sub>); optical yield, 18 %. Since the compound (<u>1</u>) was obtained by acetylation of (<u>2</u>) with acetic anhydride in pyridine<sup>3</sup>, the absolute configuration of C-5 in <u>1</u> and <u>2</u> was thus established to be <u>S</u>.

Table I. LIS values of OMe and fluorine chemical shifts of  $CF_3$  for MTPA esters

| Compound | LIS <sub>OMe</sub> a) | ∆ LIS <sub>OMe</sub> b) | CF3 <sup>c)</sup> |  |
|----------|-----------------------|-------------------------|-------------------|--|
| 3        | 1.27                  |                         | 8.48              |  |
| <u>4</u> | 1.82                  | - 0.55                  | 9.08              |  |

a) determined at a molar ratio of Eu(fod)3/ester of 1.7:1 in CCl<sub>4</sub> at 60 MHz. b) The value represents the difference in two LISOMe for MTPA esters; LISOMe(3)-LISOMe(4). c) Fluorine shifts (in ppm) are downfield from external TFA in CDCl<sub>3</sub> at 56.4 MHz.

The stereochemistry of the other chiral centers at C-6, 3a and 14a were determined by relating to the configuration of C-5 as follows. The 360 MHz <sup>1</sup>H-NMF spectrum of <u>1</u>(Fig.1) gave clear separation of all of the overlapping signals observed for the protons at C-6, 5, 4, 3a, 14a and 14 positions in the 100 MHz spectrum, and exact coupling constant values between the protons at those positions were obtained by decoupling experiments as listed in Table II.<sup>7</sup> Similar coupling constants for the protons of the same positions in <u>2</u> were observed by measuring the 360 MHz <sup>1</sup>H-NMR spectrum. Therefore, both compounds (<u>1</u>) and (<u>2</u>) were suggested to have a similar conformation, especially in the neighborhood of the chiral centers at C-6, 5, 3a and 14a.

A remarkable shifts for both H-5 and H-3a were observed in going from the alcohol (2) to the acetate (1): H-5 downfield shift from 3.72 to 5.17 ppm; H-3a upfield shift from 3.52 to 3.01 ppm (Table II). The upfield shift of H-3a of 1 can be explained by the anisotropic effect of the acetoxyl group,<sup>8)</sup> and the protot on C-3a is presumed to be extremely close proximity to the acetoxyl on C-5 as depicted in Fig.2. This arrangement was supported by the observation of the

| Compound          | <u>1</u>                      | 2                          |
|-------------------|-------------------------------|----------------------------|
| H-3a              | 3.01 (m)                      | 3.52 (m)                   |
| H-4               | 1.76 (ddd,J=15.3,11.2,2.1)    |                            |
|                   | 1.82 (ddd, J=15.3, 10.4, 4.3) | 1.68 (ddd,J=15.1,11.0,3.0) |
| <b>H-</b> 5       | 5.17 (ddd,J=10.4,3.0,2.1)     | 3.72 (tt,J=10.9,2.3)       |
| H-6               | 2.35 (m)                      | 2.34 (m)                   |
| H-10              | 5.07 (brt,J=8.0)              | 5.06 (brt,J=8.0)           |
| H <b>-</b> 14     | 5.06 (brd,J=10.0)             | 5.11 (brd,J=10.0)          |
| H-14a             | 5.37 (dd,J=10.0,8.4)          | 5.32 (dd,J=10.0,8.0)       |
| Ha                | 6.22 (d,J=3.2)                | 6.21 (d,J=3.0)             |
| НЪ                | 5.48 (d,J=2.9)                | 5.52 (d,J=2.5)             |
| COCH <sub>2</sub> | 2.19 (s)                      | 2.20 (s)                   |
| 0Ac               | 2.14 (s)                      |                            |
| OH                |                               | 3.38 (d,J=10.9)            |
| C-9 Me            | 1.58 (brs) <sup>b)</sup>      | 1.63 (brs) <sup>b)</sup>   |
| C-13 Me           | 1.86 (brs) <sup>b)</sup>      | 1.76 (brs) <sup>b)</sup>   |

Table II. 360 MHz <sup>1</sup>H-NMR data<sup>a)</sup> of <u>1</u> and <u>2</u>

a) Chemical shifts are given in  $\delta$  units with tetramethylsilane as an internal standard in  $CDCl_3(J \text{ in Hz})$ . b) These assignments may be reversed.



Fig.1 360 MHz  $^{1}$ H-NMR spectra and decoupling experiments of  $\underline{1}$ 



Fig.2 Coupling constant assignments for 1 (J in Hz)

coupling constants between H-5, H-4 and H-3. in 1. Furthermore, the coupling constant between H-3a and H-14a  $(J_{3a, 14a} = 8.4 \text{ Hz})$  is consistent with cis relationship. Accordingly, from the consideration described above, the configuration of C-5, 3a and 14a in 1 were assinged as shown in Fig.2. Determination of remaining chirality at C-6 which is shown in Fig.2, was based on the vicinal coupling constant  $(J_{5,6} = 3 \text{ Hz})$  between H-5 and H-6 in  $\underline{1}$ , and also on the presence of strong IR bands ( $\mu_{max}$ 3400, 168) cm<sup>-1</sup>) due to the intramolecular hydrogen

bonding of the hydroxyl and methyl ketone in 2. This was also supported by hydrogbonding induced downfield shift<sup>10)</sup> for the carbonyl carbon of the methyl ketone in <sup>13</sup>C-NMR spectrum of 2 (211.0 ppm) from its position in 1 (208.8 ppm).<sup>3)</sup>

Thus, the stereochemistry of both compounds  $(\underline{1})$  and  $(\underline{2})$  was established to have same absolute configuration of 3aR, 14aR, 5S and 6R as depicted in 1 and 2. Acknowledgement : The authors wish to thank Mr. John F. Kozlowski, Purdue University, for the measurement of 360 MHz <sup>1</sup>H-NMR spectra. This investigation was supported in part by the National Institutes of Health Research Grant No.RR01077 from the Division of Research Resources, which is gratefully acknowledged.

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  7) Fig.1 shows a summary of decoupling experiments of <u>1</u> as follows :

- 7) Fig.1 shows a summary of decoupling experiments of <u>1</u> as follows :

| irradiated proton(5, ppm)  | observed protons and changes(J in Hz)   |
|----------------------------|---|
| H-3a (3.01)                | $H=4(1.76, ddd) \longrightarrow dd, J=15.3, 2.1; H=4(1.82, ddd) \longrightarrow dd, J=15.3, 10.4; Ha(5.48, d) \longrightarrow s:$ |
|                            | $Hb(6.22, d) \longrightarrow s; H-14a(5.37, dd) \longrightarrow d, J=10$  |
| H-5(5.17)                  | $H-4(1.76, ddd) \longrightarrow dd, J=15.3, 11.2;$  |
| ц //1 76)                  | $H-4(1.82, ddd) \longrightarrow dd, J=15.3, 4.3$  |
|                            |   |
| H-6(2.35)                  | H=5(5.1/, ddd) dd,J=10.4,2.1  |
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